

SORPTION OF LEAD FROM AQUEOUS SYSTEM USING COCOA POD HUSK BIOCHAR: KINETIC AND ISOTHERM STUDIES

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ABSTRACT

Cocoa pod husk (CPH) was pyrolyzed at 500°C to produce biochar (CPHB) for sorption of lead (Pb) from aqueous system. Chemical characterization for CPHB was conducted using Fourier transform infrared (FTIR) spectroscopy, Boehm titration and X-ray fluorescence (XRF) spectroscopy. Sorption parameters for CPHB (i.e., sorbent dosage, pH, contact time, and input Pb concentration) were optimized. Elemental compositions for CPHB are C (66%w/w), O (19%w/w), and N (2%w/w). The ash content of CPHB consists of calcium oxide (CaO) (4.6%w/w) and potassium oxide (K₂O) (4.2%w/w), with negligible content of heavy metals (1%w/w). Upon treatment with artificial Pb wastewater, FTIR spectra for CPHB revealed shifting of *nasymm*(COO⁻) and *nsymm*(COO⁻) bands from 1560 cm⁻¹ to 1575 cm⁻¹ and 1416 cm⁻¹ to 1398 cm⁻¹, respectively. The optimum sorption parameters were determined (i.e., sorbent dosage: 1.0 g/L; pH 5; input Pb concentration; 50 mg/L; and sorption time: 210 minutes). Sorption of Pb by CPHB was best described by pseudo-second-order kinetic model ($R^2=0.835$), and Langmuir isotherm model ($R^2=0.962$). The maximum Langmuir Pb sorption capacity for CPHB (q_{max}) was 69.9 mg/g. Sorption of Pb by CPHB may have occurred through (1) coordination with polar groups (i.e., carboxylate and phenolate) and (2) precipitation with alkaline materials (i.e., CaO and K₂O).

Keywords: Carbon, pyrolysis; complexation; heavy metals; solid waste management

INTRODUCTION

Groundwater in Malaysia is contaminated with toxic lead (Pb) [1]. Exposure of Pb in groundwater may cause damage to kidney, nervous system and reproductive systems. Conventional treatment that consists of physical and chemical processes may be effective in reducing Pb concentration groundwater or leachate. However, large scale consumption of energy and chemicals increase the cost of treatment [2]. Biosorption may offers a cheaper solution for effective removal of trace Pb from groundwater. Cheap and abundant biomass from the agricultural sector may be reutilized as biosorbent for

decontamination of Pb in groundwater [3-5].

Malaysia produces large amount of agricultural biomass from cocoa plantation. The cocoa pod husk (CPH) that accounts for 70 - 75% of the total weight of the whole cocoa fruit [6], has no commercial value and is usually disposed in landfill. Recently, CPH has shown potential in decontaminating heavy metals (i.e., zinc) in wastewater [7]. The sorption capacity of CPH for heavy metals may be enhanced further when pyrolyzed at anoxic condition, whereby, dehydration of sap cells in CPH produces a carbon- rich and porous material called cocoa pod husk biochar (CPHB). In fact, biochar prepared at high temperature (i.e., >500°C) is more stable [8], and may be suited for decontamination of Pb from aqueous system.

To the best of our knowledge, Pb sorption kinetics and isotherms for CPHB has never been reported. This study aims to investigate the potentials of CPHB as sorbent for Pb, by measuring maximum Pb sorption capacity from artificial groundwater, and to determine the Pb sorption mechanism of CPHB.

EXPERIMENTAL

Synthesis of CPHB

Cocoa pod husk (CPH) was collected from the Malaysian Cocoa Board Centre of Research and Development, Jengka, Pahang, Malaysia. The as-received CPH was rinsed with deionized water (DI) to remove impurities. Then, CPH was oven-dried at 60°C for 72 hours. The dried CPH was pulverized with a hammer-mill machine, and sieved with a 70 mesh screen to 200 µm in particle diameter. The fine CPH particles was pyrolyzed in a furnace at 500°C under nitrogen gas ambient for 2 hours to produce CPHB. The CPHB was washed thoroughly with DI water, oven-dried at 60°C for 72 hours, and homogenized for further analyses.

Characterization for CPHB

The pH for CPHB was determined from 10 mmol/L calcium chloride (CaCl₂) solution (CPHB/solution ratio 1:10) using a calibrated pH meter. The elemental analyses was conducted at the Institute of Science, UiTM Shah Alam, using elemental analyser (ThermoFinnigan Flash EA2000, UK), and XRF spectroscopy (PANalytical Epsilon3-XL, Netherlands). The FTIR spectra was recorded using FTIR spectrometer (Perkin Elmer Spectrum One, USA) on KBr disc containing 5% w/w of samples. The average FTIR spectra for CPHB and Pb-treated CPHB were determined from 64 scans between 4000 cm⁻¹ and 400 cm⁻¹ wavenumber. Boehm titration was conducted according method described by Goertzen *et al.* (2010). The amount of functional groups on the surface of CPHB were calculated with Eq 1 [9]:

$$\text{mol carbon functionality} = \frac{([R_b] \times V_r) - ([X] \times V_x) - ([T] \times V_t)}{m} \quad (1)$$

where [R_b] is the concentration of base reacted with CPHB (mmol/L); V_r is the volume

of base reacted with CPHB (L); [X] is the concentration of excess hydrochloric acid added (mmol/L); V_x is the volume of excess hydrochloric acid added (L); [T] is the concentration of NaOH titrant (mmol/L); V_t is the volume of NaOH titrant (L) and m is mass of CPHB used in Boehm titration (g).

Batch sorption experiments.

Sorption experiment was conducted using 50 mg/L Pb nitrate solution as synthetic wastewater. Potassium nitrate (10 mmol/L) was added to lead nitrate solution as background electrolyte for maintaining ionic strength of Pb nitrate solution. Sorption experiment was optimized at five sorbent dosage values (i.e., 0.05, 0.10, 0.50, 1.0, 5.0 g/L), and five solution pH values (i.e., 3, 4, 5, 6, 7). Sorption kinetic was conducted at optimized solution pH and sorbent dosage. Nineteen data points were collected between 0 and 210 min of contact time. For sorption isotherm experiment, CPHB was contacted with five initial Pb concentrations (i.e., 10, 20, 30, 40, and 50 mg/L) at optimized solution pH, sorbent dosage and contact time. The mixtures were filtered with filter paper to separate spent CPHB. The residual Pb concentrations of the filtrates were analyzed using flame atomic absorption spectroscopy (FAAS) (Perkin Elmer AAnalyst 400, USA). The sorption capacity (q_t) was calculated with Eq 2:

$$q_t = \frac{(C_o - C_t)V}{m} \quad (2)$$

where C_o and C_t is the initial and equilibrium Pb concentration (mg/L), respectively; V is the volume of residual Pb (L); and m is the dry weight of CPHB (g). The sorption data was analyzed with kinetic models (i.e., pseudo-first-order and pseudo-second-order) and isotherm models (i.e., Langmuir and Freundlich) (Table 1) [10].

Table 1: Non-linear kinetic models (pseudo-first-order & pseudo-second-order) and isotherm models (Langmuir & Freundlich)

Kinetic/isotherm models	Non-linear equation
Pseudo-first-order	$q_t = q_e(1 - \exp(-k_1 t))$
Pseudo-second-order	$q_t = \frac{C_e^2 k_2 t}{1 + C_e k_2 t}$
Langmuir	$q_t = \frac{q_{\max} K_L C_e}{1 + K_L C_e}$
Freundlich	$q_t = K_F C_e^{1/n}$

where q_e (mg/g) is the amount of the metal ions adsorbed at equilibrium (mg/g); t (minutes) is the contact time for CPHB and Pb nitrate solution; k_1 (min^{-1}) and k_2 (g/mg min) are the rate constant for the pseudo-first-order and pseudo-second-order equation, respectively; q_{max} is the maximum Langmuir sorption capacity of CPHB (mg/g); K_L is the Langmuir constant (L/mg); K_F is the Freundlich constant (mg/g); and n is the empirical constant (L/mg). The sorption data were fitted with non-linear kinetic and isotherm models using Sigmaplot 11.0 software.

RESULTS AND DISCUSSION

Characterization for CPHB

Elemental compositions for CPHB are shown in Table 2 (i). Cocoa pod husk biochar primarily consists of C (65.7%w/w), with 10.1%w/w of ash content. Further XRF analysis shows that the ash in CPHB constitutes CaO (4.6%w/w) and K₂O (4.2%w/w), and with negligible contents of CuO (0.03%w/w), ZnO (0.19%w/w) and MnO (0.46%w/w). Both CaO and K₂O are alkaline materials and may have contributed to the high pH (9.1 ± 0.32) of the CPHB in CaCl₂ solution. Based on the results from Boehm titration (Table 2 (ii)), the contents for phenolic and carboxylic groups on the surface of the CPHB are 0.75 mmol/g and 1.5 mmol/g, respectively.

Table 2: (i) Elemental and ash composition [%w/w] and (ii) functional groups composition [mmol/g] in CPHB

(i)		(ii)	
Elements	Composition [%w/w]	Functional groups	Composition [mmol/g]
C	65.7	Carboxylic	1.5
H	2.7	Lactonic	0
N	2.2	Phenolic	0.75
O	19.3	Surface acidity	2.25
Ash	10.1	Surface basicity	0.66

The wavenumber values for bands in the FTIR spectra for CPHB and the Pb-CPHB are shown in Table 3. The presence of phenolic and carboxyl groups in Boehm titration experiment corroborates with the FTIR result. The strong $\nu(\text{COO}^-)$ vibration band in the FTIR spectra CPHB were observed at 1575– 1690 cm^{-1} . Upon treatment with Pb nitrate solution, the $\nu_{\text{asym}}(\text{C-O})$ and $\nu_{\text{sym}}(\text{C-O})$ vibration bands of the carboxylate group shifted from 1560 cm^{-1} to 1575 cm^{-1} , and 1416 cm^{-1} to 1398 cm^{-1} , respectively. The $\Delta\nu(\text{COO}^-)$ value [$\nu_{\text{asym}}(\text{COO}^-) - \nu_{\text{sym}}(\text{COO}^-)$] in the Pb-CPHB FTIR spectra is 177 cm^{-1} , indicating that Pb(II) ion chemically bonded to the carboxylate group through monodentate coordination (figure 1) [11]. A shoulder at 1243 cm^{-1} in Pb-CPHB spectra was assigned to stretching vibration for phenolate group [12]. Two stretching bands of aliphatic hydroxyl groups in CPHB (at 1053 cm^{-1} and 1043 cm^{-1} , respectively [13]) remained similar even after treatment with Pb nitrate solution. Therefore, coordination of Pb(II) ions did not occur with the hydroxyl groups in CPHB.

Table 3: FTIR bands wavenumbers [cm^{-1}] for CPHB and Pb-CPHB spectra

Samples	Band wavenumber [cm^{-1}]			
	$\nu_{\text{asymm}}(\text{COO}^-)$	$\nu_{\text{symm}}(\text{COO}^-)$	$\nu(\text{CH-OH})$	$\nu(\text{CH}_2\text{-OH})$
CPHB	1560m	1416m	1053vs	1043vs
Pb-CPHB	1575vs	1398vs	1053vs	1045vs

Note: Peak intensity (with respect to the strongest peak in the spectrum): vs (very strong) 80–100%, s (strong) 60–80%, m (medium) 40–60%, w (weak) 20–40%, vw (very weak) 0–20%; Bond vibration mode: ν = stretching, sym = symmetric, asymm = asymmetric



Figure 1: Monodentate coordination of Pb(II) ion with the carboxylate group in CPHB

Batch sorption experiments

The q_t values [mg/g] as a function of initial solution pH and sorbent- solution ratio [g/L] are shown in figure 2. The q_t values for CPHB is increased with increasing sorbent solution ratio and solution pH. The q_t values reached plateau when sorbent-solution dosage is 1.0 g/L , and solution pH is around 6-7. Sorption for Pb were enhanced possibly due to (1) stronger interaction between Pb(II) ions and deprotonated carboxyl group in CPHB, and (2) minor precipitation of Pb(II) ions to insoluble lead hydroxide, carbonate, phosphate or sulfate species [10]. The regression plots for non- linear kinetic and isotherm models are shown in figure 3. Sorption kinetic parameters were shown in Table.

Sorption data has high fitness to the pseudo-second-order kinetic model ($R^2 = 0.835$), showing that the rate limiting step may be controlled by chemical process [14]. Sorption isotherm parameters were shown in Table 5. Sorption of Pb by CPHB is best described by the linearized Langmuir isotherm model ($R^2 = 0.962$), indicating the Pb monolayer adsorbed on the homogenous surface of CPHB. The Langmuir constant (RL) was 0.091-0.33, indicating that Pb sorption is a favourable process [14].

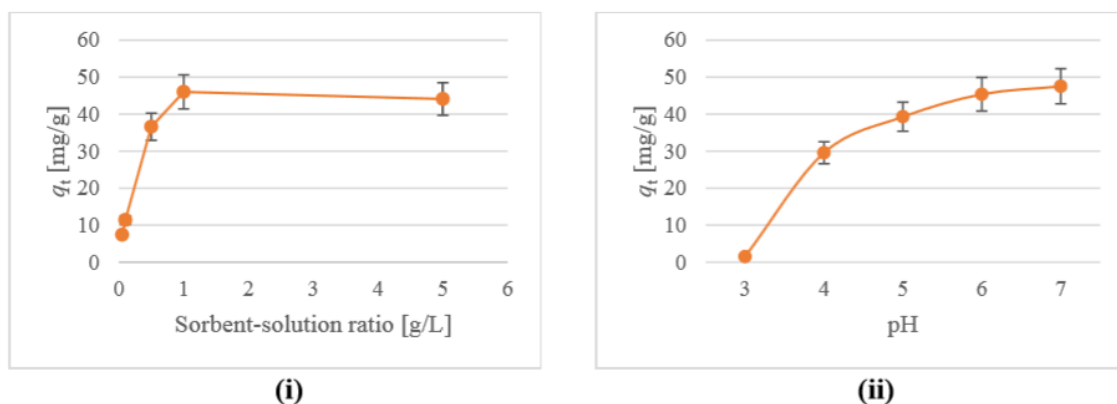


Figure 2: Lead sorption capacity for CPHB (q_t , mg/g) as a function of (i) sorbent/solution ratio (g/L), and (ii) solution pH

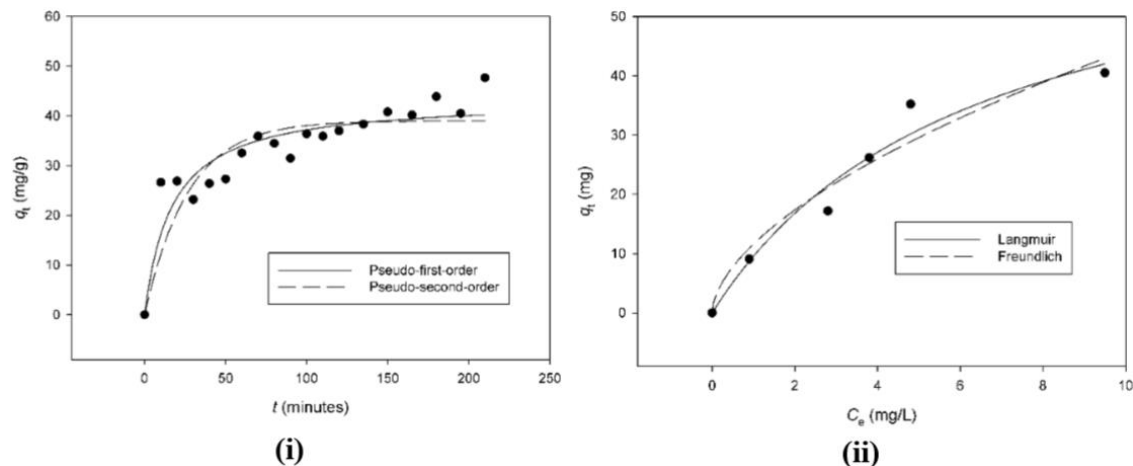


Figure 3: Non-linear regression plots for (i) kinetic models (pseudo-first-order & pseudo-second-order); and (ii) isotherm models (Langmuir & Freundlich)

Table 4: Kinetic models parameters for Pb sorption by CPHB

Kinetic models	R^2	q_e [mg/g]	k
Pseudo-first-order	0.753	39.0	0.0373
Pseudo-second-order	0.835	43.5	0.0014

Table 5: Isotherm models parameters for Pb sorption by CPHB

Isotherm models	R^2	$q_{\max}$ [mg/g]	K_L [mg/L]	K_f [mg/g]	n
Langmuir	0.962	69.9	0.158	-	-
Freundlich	0.946	-	-	11.6	1.720

The $q_{\max}$ value for CPHB (69.9 mg/g) is significantly higher than those of cocoa pod powder (5.31 mg/g) [15] and cocoa shell (6.54 mg/g) [16]. As a comparison with other biomass-based sorbent available in Malaysia, the $q_{\max}$ value for CPHB is lower than palm shell activated carbon (95.2 mg/g) [17], but is higher than bone powder (55.3 mg/g), active carbon (41 mg/g), and commercial carbon (27.3 mg/g) [18].

CONCLUSION

Pyrolysis of CPH has produced CPHB with a high Pb sorption capacity that is comparable to commercial sorbents available in Malaysia. Lead ions monolayer may adsorbed on the CPHB surface through monodentate coordination bonding with carboxylate or phenolate groups. Possible precipitation of Pb(II) ions in the aqueous phase by CaO and K₂O may have contributed to Pb sorption capacity for CPHB.

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